

Double mismatch interactions in probe design enable improved hybridization-based SNV discrimination

Thijs Van der Snickt¹, Yannick Stulens², Laura Claudino¹, Simone Bassini^{3,4}, Scott Ailliet¹
 Karolien De Wael^{3,4}, Jef Hooyberghs², Pieter Mestdagh¹

¹OncoRNALab, Department of Biomolecular Medicine, Ghent University, 9000, Ghent, Belgium

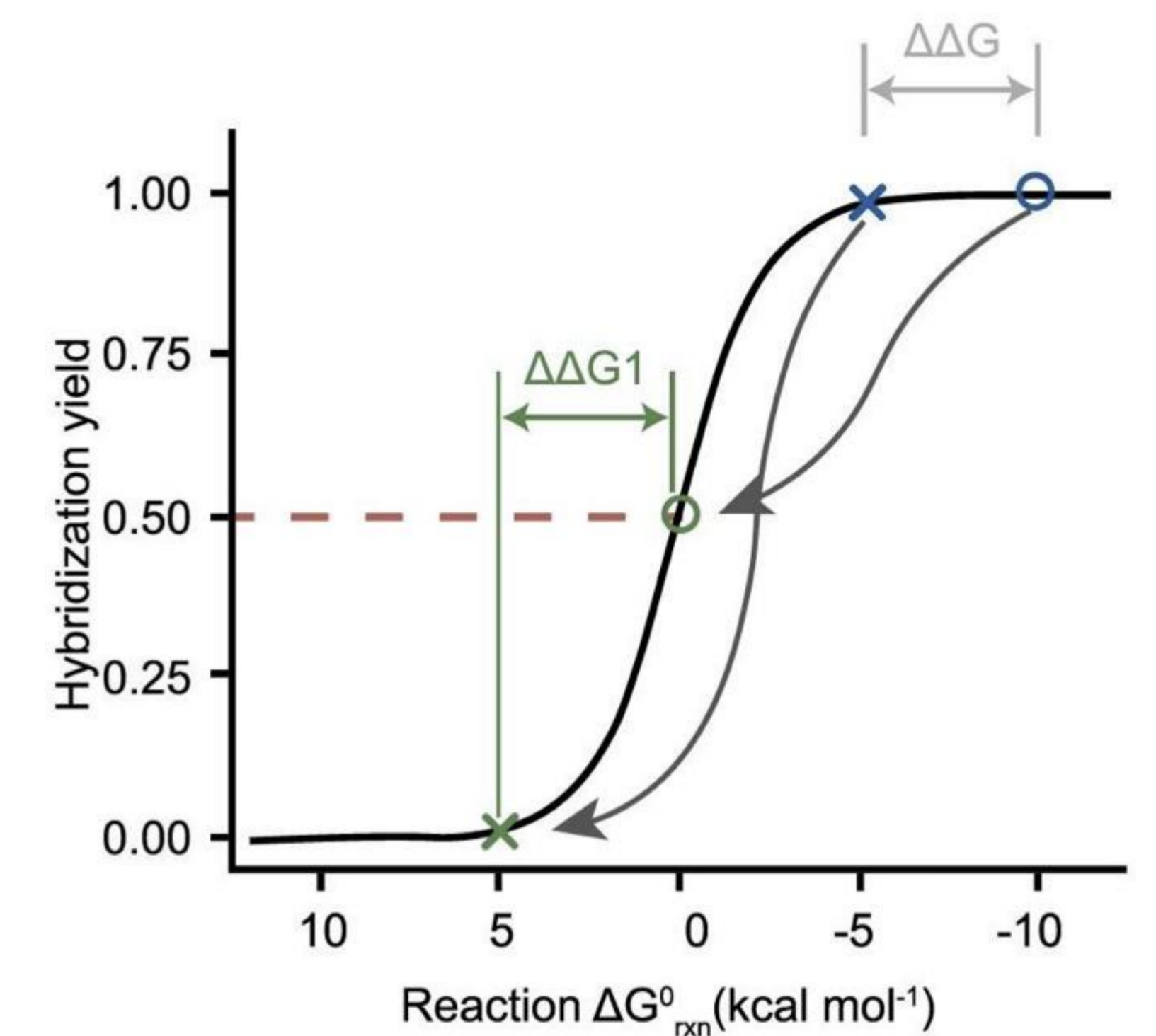
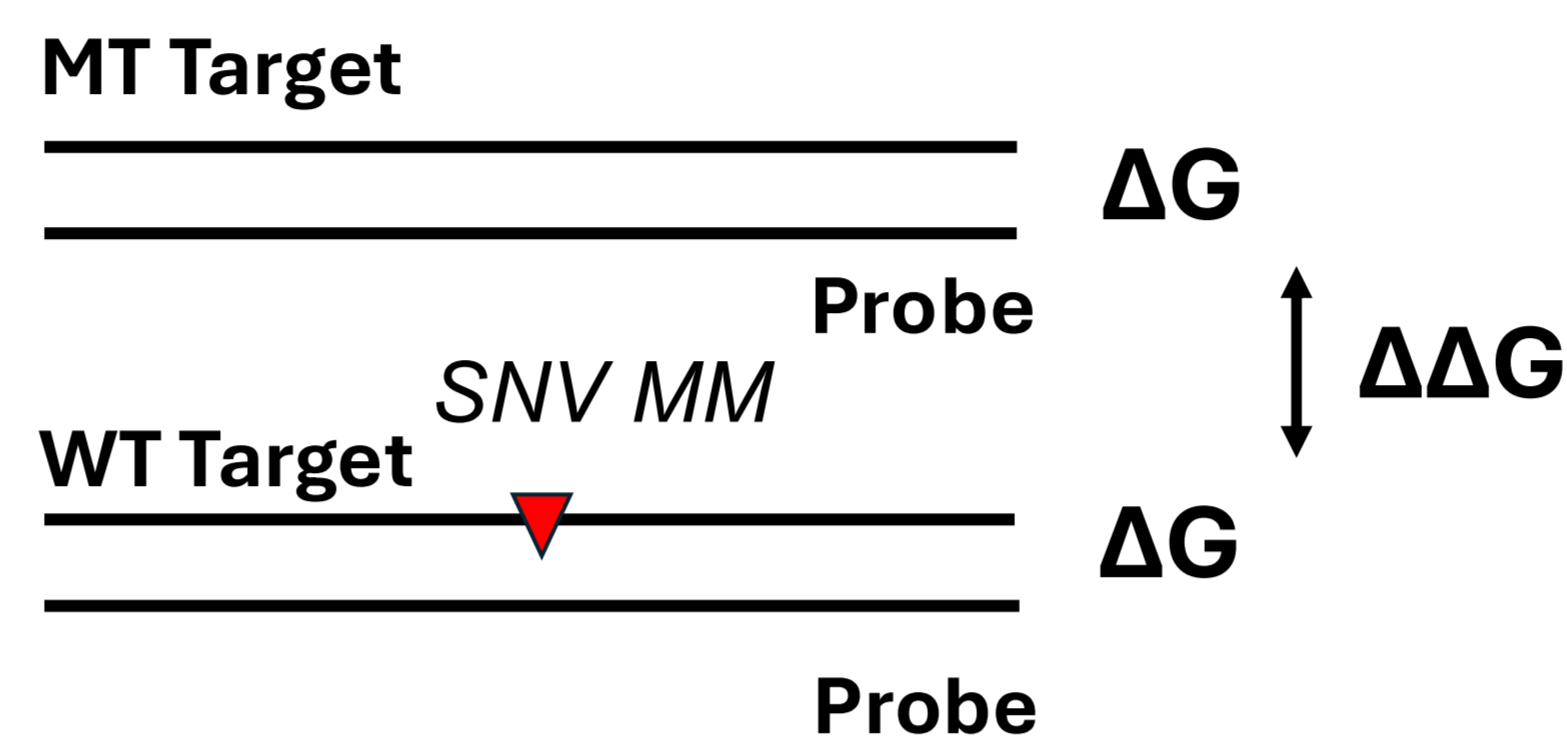
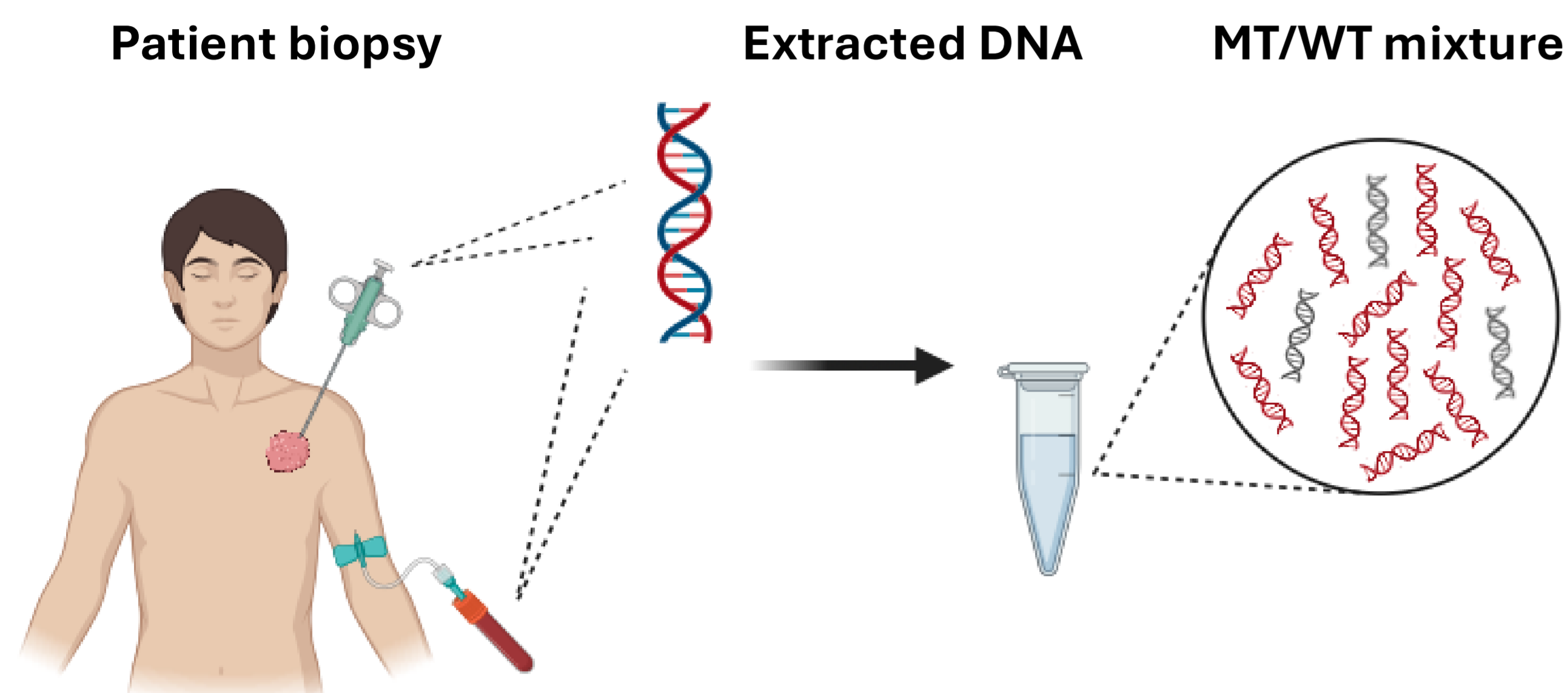
³Photoelectrochemistry and sensing (A-PECS), University of Antwerp, 2020, Antwerp, Belgium

²UHasselt, Data Science Institute, Theory Lab, Agoralaan, 3590, Diepenbeek, Belgium

⁴NANOlight Center of Excellence, University of Antwerp, 2020, Antwerp, Belgium

INTRODUCTION

MT and WT sequences differ by only one nucleotide, leading to nearly identical probe binding energy (ΔG)

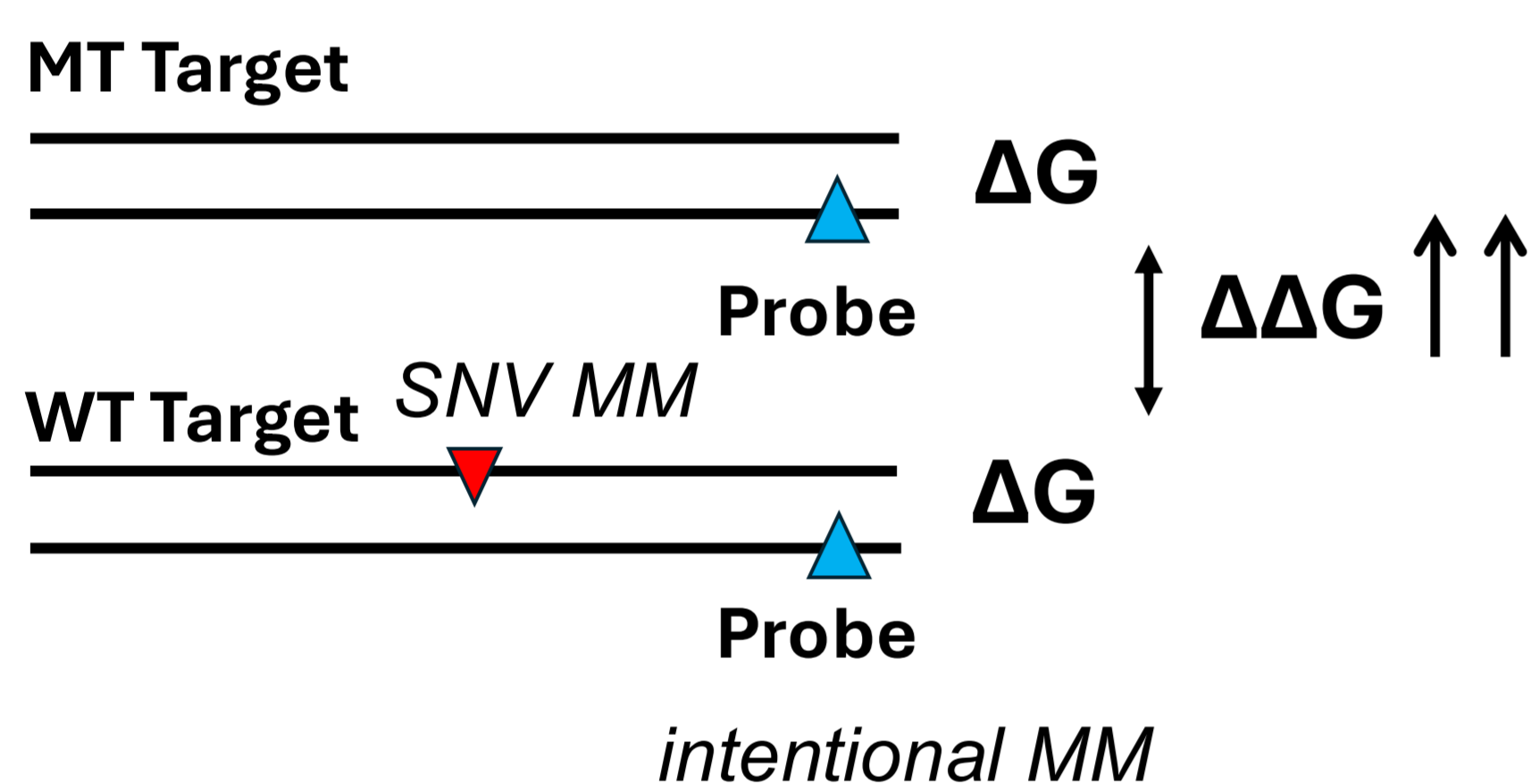


Single-Nucleotide Variants (SNVs) differ from wild-type (WT) DNA by only one base (red triangle), leading to nearly identical probe binding affinities (ΔG). This high homology causes rare mutant (MT) signals to be overwhelmed by WT background. Classical "equilibrium shifting methods" (e.g., higher hybridization temperature, shorter probes, or LNAs/PNAs) increases stringency but shifts both hybridization curves simultaneously. While this reduces unintended WT binding (X), it causes a proportional loss in MT signal (O). Because intrinsic thermodynamic gap ($\Delta\Delta G$) remains constant, gaining selectivity requires a sacrifice in sensitivity.

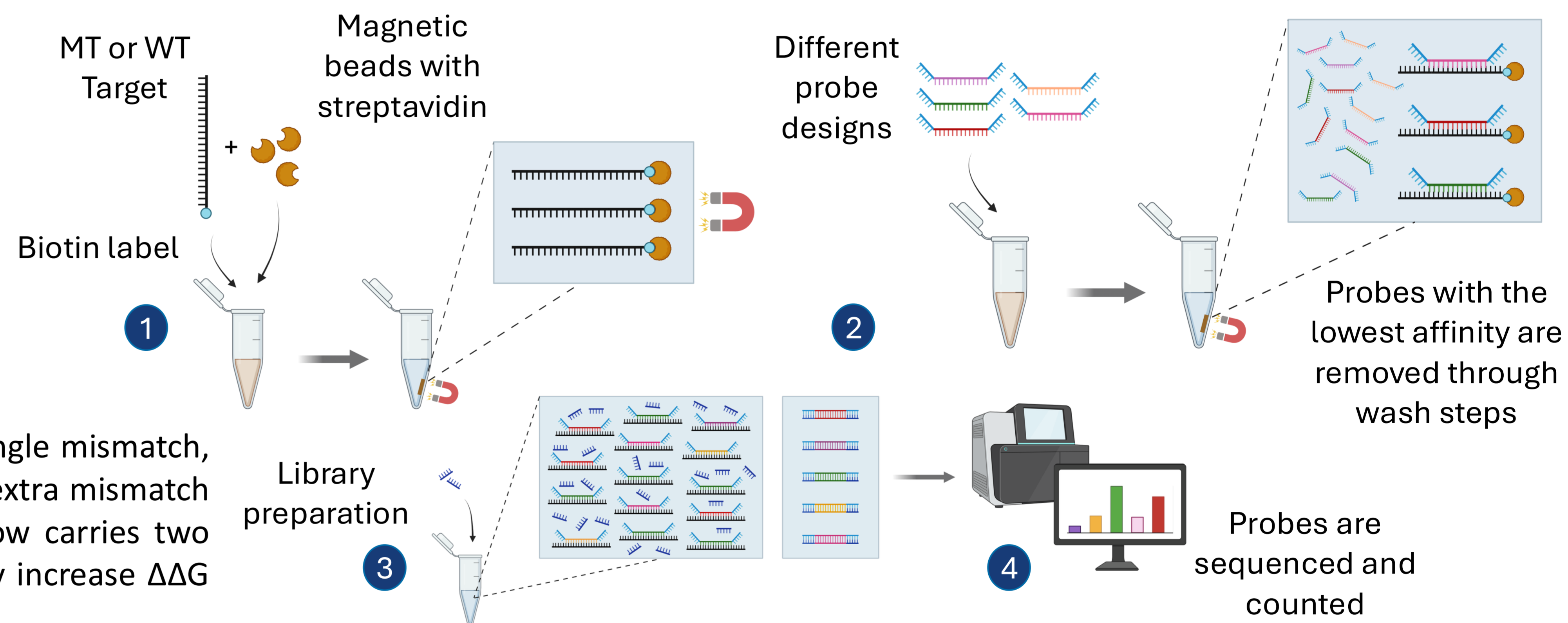
METHODS

Intentional mismatches can destabilize wild-type binding

Probe enrichment platform was developed to investigate probe hybridization



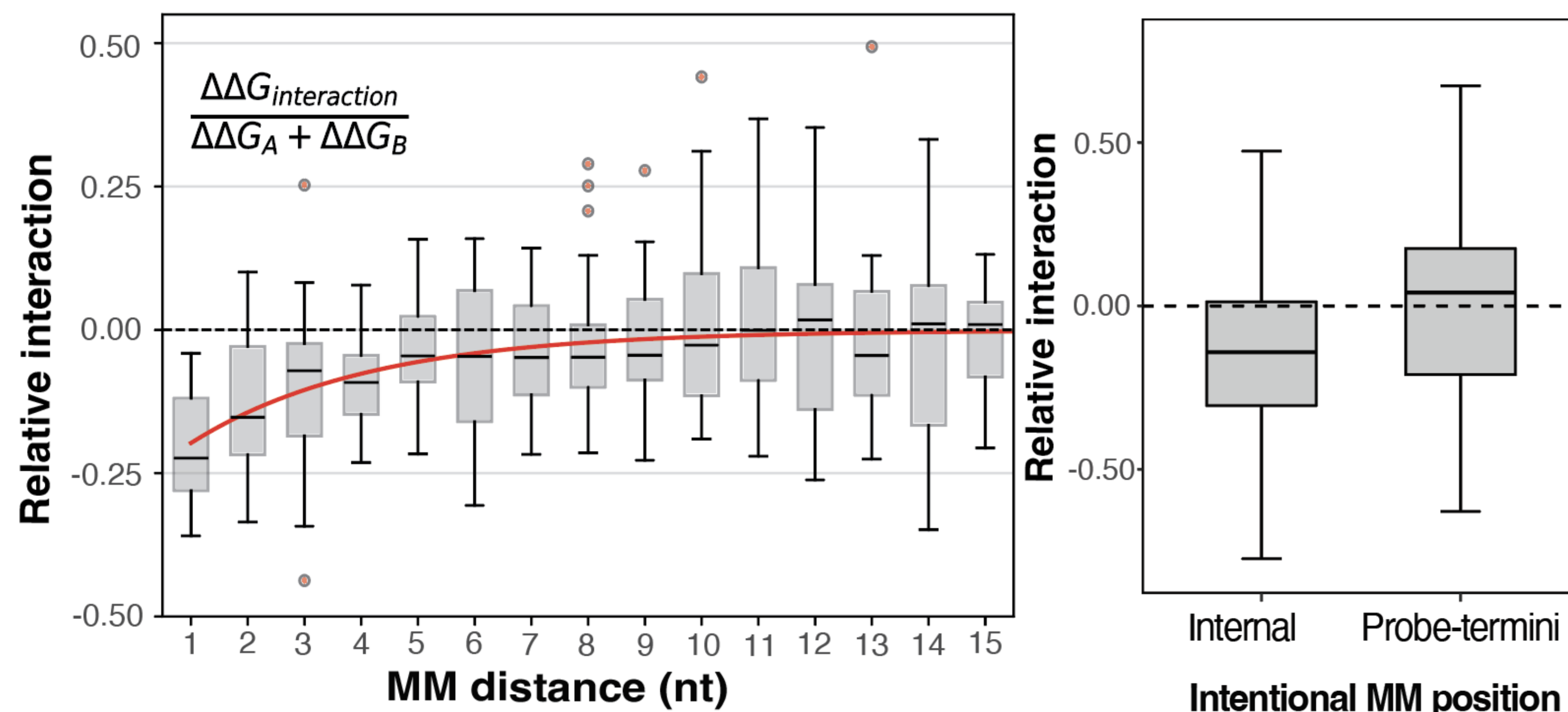
In classic probe design, selectivity is limited by the small $\Delta\Delta G$ of a single mismatch, making it difficult to distinguish MT from excess WT. Introducing an extra mismatch adds an additional free-energy penalty, especially for WT, which now carries two mismatches. If these mismatches interact in a destabilizing way, they increase $\Delta\Delta G$ beyond the single-mismatch limit, leading to higher selectivity.



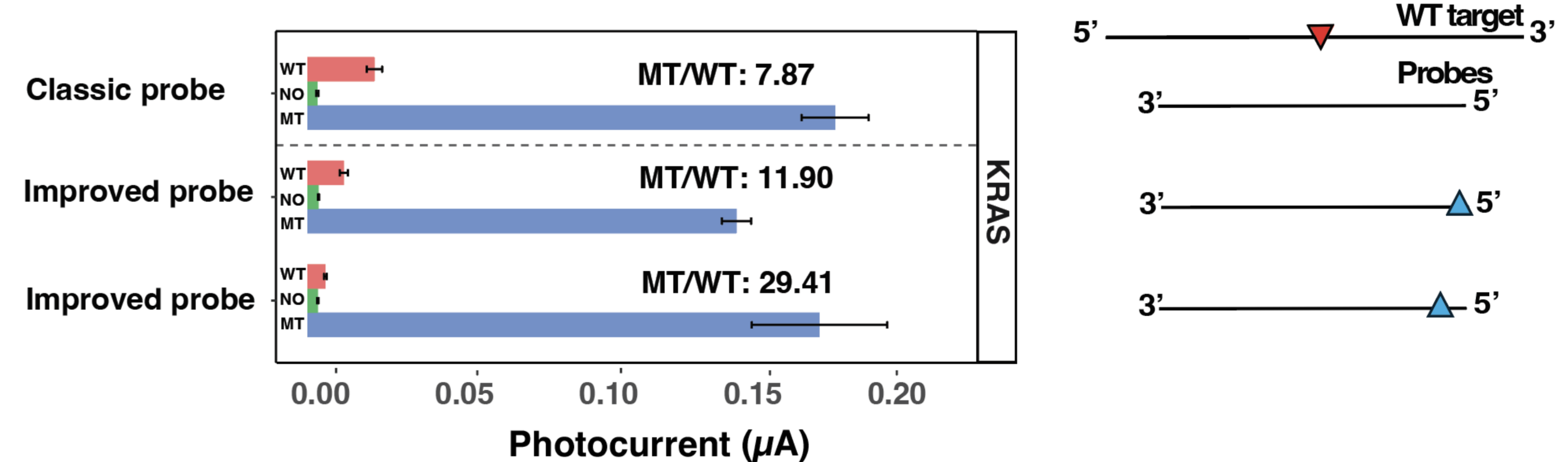
RESULTS

Mismatch position and interaction determine hybridization selectivity

Probe screening for KRAS p.G12C and BRAF p.V600E revealed improved probe designs that increased $\Delta\Delta G$



While each mismatch (MM1 ($\Delta\Delta G_A$) or MM2 ($\Delta\Delta G_B$)) contributes a fixed free-energy penalty, combining them introduces an interaction term ($\Delta\Delta G_{int}$) that alters duplex stability beyond simple addition. Closely spaced mismatches tend to stabilize the duplex, whereas more distant mismatches can cause additional destabilization. This increases the total $\Delta\Delta G$ beyond additive effects, improving target discrimination.



Systematic screening of 884 probe variants for KRAS p.G12C and BRAF p.V600E reveals that "classic" perfectly matched probes are often outperformed by rationally designed alternatives. By pairing a central variant with an intentional terminal mismatch, the $\Delta\Delta G$ is maximized. When benchmarked on a PEC platform, this design motif achieved a 29.41 fold MT/WT (for KRAS p. G12C) selectivity ratio, a nearly 4-fold improvement over classic designs.