



Comprehensive benchmarking of computational deconvolution of transcriptomics data

Francisco Avila Cobos

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Who are we?



HOME

ABOUT US

PROFESSIONALS

PATIENT

RESEARCH





www.cmgg.be/en/



www.crig.ugent.be

NEUROBLASTOMA

BIOINFORMATICS

THE NON-CODING TRANSCRIPTOME

DATA-MINING

PAN-CANCER

Who are we?



Garvan Institute of Medical Research

Single Cell and Computational Genomics



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Even though scientific research should guarantee reproducibility and replication of any experiment...

- Neuroblastoma
- ALL, AML, ...
- Lung cancer
- Cancer 'X'



There is a clear "reproducibility crisis" in scientific research



There is a clear "reproducibility crisis" in scientific research



1,500 scientists lift the lid on reproducibility. Baker, M. (2016) Nat. News, 533,452.

There are several important factors responsible for this crisis

Sample heterogeneity

Insufficiently documented or incorrect data processing practices

(MAQC Consortium, 2010).

Platform-specific differences (SEQC/MAQC-III Consortium, 2014).

Tumor samples also contain a variable portion of non-malignant cells that include epithelial, stromal and infiltrating immune cells



Tumors as organs: complex tissues that interface with the entire organism. Egeblad et al., 2010. *Dev Cell*. 18(6):884-901.

Single-cell technologies allow the analysis of individual cells within heterogeneous tissues...



...but have labourintensive protocols and require expensive resources, hindering its establishment in the clinic



Single-cell technology is not applicable to cell-free scenarios



Integrating liquid biopsies into the management of cancer. Siravegna et al., 2017. Nature Reviews Clinical Oncology

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Charting extracellular transcriptomes in The Human Biofluid RNA Atlas

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Eva Hulstaert





Lucía Lorenzi

The RNA Atlas, a single nucleotide resolution map of the human transcriptome

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 Pieter Mestdagh



small RNA sequencing polyA RNA sequencing total RNA sequencing Computational deconvolution is the solution



DECONVOLUTION inference of cell type proportions AND/OR cell type-specific expression profiles

in heterogeneous samples

Deconvolution applied to cell-free scenarios has been mainly focused on DNA



CancerLocator: non-invasive cancer diagnosis and tissue-of-origin prediction using methylation profiles of cell-free DNA

nature genetics

Identification of methylation haplotype blocks aids in deconvolution of heterogeneous tissue samples and tumor tissue-of-origin mapping from plasma DNA



Although deconvolution of cfRNA also exists



National Academy of Sciences of the United States of America

Noninvasive in vivo monitoring of tissuespecific global gene expression in humans



Winston Koh, Wenying Pan, Charles Gawad, H. Christina Fan, Geoffrey A. Kerchner, Tony Wyss-Coray, Yair J. Blumenfeld, Yasser Y. El-Sayed, and Stephen R. Quake



Bioinformatics

Computational deconvolution of transcriptomics data from mixed cell populations

Francisco Avila Cobos^{1,2,3}, Jo Vandesompele^{1,2,3}, Pieter Mestdagh^{1,2,3,†} and Katleen De Preter^{1,2,3,*,†}



- Mathematical approaches
- Factors affecting the deconvolution efficiency:
 - Pre-processing

. . .

• Logarithmic versus linear space

Although log transformation is routinely included as part of the pre-processing of omics data...

Expression data often transformed into logarithmic scale because the **statistical tests used** for differential gene expression assume an underlying normal distribution.



...deconvolution requires data in linear scale as opposed to log transformations for DGEA



- The reconstructed signal is an underestimation of the signal measured from the mixture.
- If the data was transformed back to linear scale → accurate deconvolution

Gene expression deconvolution in linear space. Zhong, Y. and Liu, Z. (2012) Nat. Methods, 9, 8–9.

...deconvolution requires data in linear scale as opposed to log transformations for DGEA

Deconvolution is modeled by a linear equation $\mathbf{O} = \mathbf{S} \times \mathbf{W}$, where \mathbf{O} is the expression data for mixed tissue samples, **S** is the tissue-specific expression profile, and **W** is the cell-type frequency matrix. If the signal is logtransformed, the linearity will no longer be preserved. The concavity feature of the log function will induce a downward bias to the reconstructed signal (Fig. 1a and Supplementary Fig. 1). Mathematically, it can be shown that the deconvolution model used on log-transformed signals is $log(\mathbf{O}') = log(\mathbf{S}) \times \mathbf{W}$, where \mathbf{O}' is the csSAM estimate of geneexpression profiles. As **W** is a frequency matrix and its column values sum to 1, the following is true by the properties of concave functions³: $\log(\mathbf{S} \times$ W) > log(S) × W. Taking these two equations together, we can conclude that $\log(\mathbf{O}') < \log(\mathbf{S} \times \mathbf{W}) = \log(\mathbf{O})$. Thus, we proved that when log-transformed signal is used as the input for signal reconstruction, it will always yield an underestimation of the true signal. By taking an anti-log transformation, we obtained an unbiased reconstruction of the mixed tissue samples (Fig. 1b and Supplementary Fig. 2).

Gene expression deconvolution in linear space. Zhong, Y. and Liu, Z. (2012) Nat. Methods, 9, 8–9.

Comprehensivebenchmarkingofcomputationaldeconvolution of transcriptomics data

- What's more important: transformation, pre-processing, method?
- Are they equally important?
- Are there differences in terms of performance?
- Pre-print will be available @ bioRxiv on December 5, 2019

Goal: inference of cell type proportions in artificial tissues



We take advantage of having individual cells (scRNA-seq)



single cells from different individuals

The performance is assessed using pearson correlation and the root mean squared error (RMSE)



Scenario 1: Computational deconvolution using "bulk" RNA-seq data



Avila Cobos et al., in preparation

- Supervised:
 - a) Given T and C \rightarrow P
 - OLS, nnls, RLR, FARDEEP, CIBERSORT, **MMAD**, DSA
 - b) Given T and P \rightarrow C
 - LRCDE, MMAD



BULK

- Supervised:
 - a) Given T and C \rightarrow P
 - OLS, nnls, RLR, FARDEEP, CIBERSORT, MMAD
 - b) Given T and P \rightarrow C
 - LRCDE, MMAD

BULK

- Semi-supervised: Given T + set of markers → P
 - DSA, ssKL, ssFrobenius
 - WISP (NNLS).
- Unsupervised (=Complete deconvolution): Given T \rightarrow C and P
 - MMAD, deconf, NMF (Virtual microdissection)
 - deconICA



$$\mathbf{T} = \mathbf{C} \cdot \mathbf{P}$$

 $y = X\beta$

$$\min_{P(\text{or }C)} ||C \cdot P - T||^2$$

$$\bigcirc$$

$$OLS: RSS(\beta) = (\mathbf{y} - \mathbf{X}\beta)^{\mathsf{T}}(\mathbf{y} - \mathbf{X}\beta)$$

$$\hat{\beta} = (X'X)^{-1}X'y$$

- NNLS (non-negative least squares):
 - OLS + non-negativity + sum-to-one
- RLR/FARDEEP (Robust Linear Regression):
 - Outlier removal before coefficient estimation



$$\mathbf{T} = \mathbf{C} \cdot \mathbf{P}$$

$$y = X\beta$$

$$\min_{P(or C)} ||C \cdot P - T||^2$$

NMF

- Random initializations of P
- ssNMF (ssKL, ssFrobenius)
 - Use marker information
- DSA:

$$\begin{pmatrix} g_{11} & 0 & \dots & 0 \\ g_{21} & 0 & \dots & 0 \\ 0 & g_{32} & \dots & 0 \\ 0 & g_{42} & \dots & 0 \\ 0 & g_{52} & \dots & 0 \\ 0 & 0 & \ddots & \vdots \\ 0 & 0 & \dots & g_{mk} \end{pmatrix} \Rightarrow \begin{pmatrix} \bar{g}_1 & 0 & \dots & 0 \\ 0 & \bar{g}_2 & \dots & 0 \\ 0 & 0 & \ddots & 0 \\ 0 & 0 & \dots & \bar{g}_k \end{pmatrix} \longrightarrow OLS$$



$$\mathbf{T} = \mathbf{C} \cdot \mathbf{P}$$

$$y = X\beta$$

$$min_{P(or C)}||C \cdot P - T||^2$$

$$\frac{1}{2} \|Qx - c\|^2 = \frac{1}{2} (Qx - c)^T (Qx - c) = \frac{1}{2} \left(x^T Q^T Qx - 2x^T Q^T c + c^T c \right)$$
$$\frac{1}{2} x^T Ax + q^T x$$

Regularization (lasso, ridge, elastic net)

$$\tilde{\beta} = \underset{\beta}{\operatorname{argmin}} \left\{ \sum_{i=1}^{N} \left(y_i - \beta_0 - \sum_{j=1}^{p} x_{ij} \beta_j \right)^2 + \lambda \sum_{j=1}^{p} |\beta_j|^q \right\}$$

Hastie et al. - The Elements of Statistical Learning (book)

• **WISP** (Weighted In Silico Pathology; Blum *et al.*, 2019):

Transcriptomics or methylation data

Two-step approach:

- 1) Estimates pure population profiles based on predefined pure samples.
- 2) Estimates the proportions of each pure population in a mixed sample through NNLS.

MPM **signatures** with **three components:** <u>epithelioid-like</u>, <u>sarcomatoid-like</u> and <u>non-tumor</u>.

- a) for tissues (with a complex microenvironment present)
- b) for cell lines

- deconICA (Czerwinska *et al.*):
 - Challenge: assigning components to specific biological processes, cell types and technical factors.
 - Cell-types associated with components through highest correlation.

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Cluster	Compo- nent	Meaning
Immune	RIC2	B cells
	RIC25	T cells
	RIC27	B cells
	RIC28	response to wounding
	RIC37	IFN signalling pathway
	RIC57	monocytes

Skin-related	RIC5	epidermis development and
		keratinisation
	RIC7	epidermis development and
		keratinisation
	RIC19	epidermis development and
		keratinisation
	RIC31	epidermis development and
		keratinisation

Nazarov et al, 2019. BMC Medical Genomics

Data transformation has a dramatic impact on the deconvolution results



Different combinations of normalization and method reveal important differences in performance



Removing cell types from the reference matrix results in substantially worse deconvolution results



Human pancreas

Scenario 2: Computational deconvolution using scRNA-seq data



Avila Cobos et al., in preparation

SINGLE-	
CELL	

- Supervised:
 - a) Given T and C \rightarrow P
 - DeconvSeq, MuSiC, SCDC, DWLS,...

MuSiC: MUlti-Subject Single Cell deconvolution



- DWLS (Tsoucas *et al.*, 2019): w-NNLS tweaked to adjust the contribution of each gene (e.g. <u>avoid minimal contribution of good markers only due</u> to low mean expression levels).
- SCDC (Dong *et al.*, 2019): w-NNLS + integrating multiple single-cell datasets at once while accounting for batch effects.



Data transformation has a dramatic impact on the deconvolution results



Take-home messages

- Logarithmic transformation results in a poor performance.
- Computational deconvolution must be performed with data in linear scale.
- Different combinations of normalization and method reveal important differences in performance.
- Single-cell methods have comparable performance to the best performing bulk methods.
- Removing cell types from the reference matrix leads to worse results in both bulk and single-cell deconvolution frameworks.
- **Further reading**: Sturm *et al*. Comprehensive evaluation of transcriptomebased cell-type quantification methods for immuno-oncology

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Questions?





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